

4. Lee QP, Juchau MR and Creech Kraft J, Microinjection of cultured rat embryos: Studies with retinol, 13-*cis*- and all-*trans*-retinoic acid. *Teratology* **44**: 313–323, 1991.
5. Creech Kraft J, Bechter R, Lee QP and Juchau MR, Microinjections of cultured rat conceptuses: Studies with 4-oxo-all-*trans*-retinoic acid, 4-oxo-13-*cis*-retinoic acid and all-*trans*-retinoyl- $\beta$ -glucuronide. *Teratology* **45**: 259–270, 1992.
6. Creech Kraft J, Bui T and Juchau MR, Elevated levels of all-*trans*-retinoic acid in cultured rat embryos 1.5 hr after intraamniotic microinjections with 13-*cis*-retinoic acid or retinol and correlations with dysmorphogenesis. *Biochem Pharmacol* **42**: R21–R24, 1991.
7. Stark KL and Juchau MR, Microinjection of cultured rat embryos: A new technique for studies in chemical dysmorphogenesis. *Toxicol Appl Pharmacol* **100**: 411–416, 1989.
8. Stark KL, Lee QP, Namkung MJ, Harris C and Juchau MR, Dysmorphogenesis elicited by microinjected acetaminophen analogs and metabolites in rat embryos cultured *in vitro*. *J Pharmacol Exp Ther* **255**: 74–82, 1990.
9. Creech Kraft J, Eckhoff Chr, Kuhn W, Loeffberg B and Nau H, Automated determination of 13-*cis*- and all-*trans*-retinoic acid, their 4-oxo-metabolites and retinol in plasma, amniotic fluid and embryo by reversed-phase high-performance liquid chromatography with a precolumn switching technique. *J Liq Chromatogr* **11**: 2051–2069, 1988.
10. Creech Kraft J and Juchau MR, Correlations between conceptual concentrations of all-*trans*-retinoic acid and dysmorphogenesis after microinjections of all-*trans*-retinoic acid, 13-*cis*-retinoic acid, all-*trans*-retinoyl- $\beta$ -glucuronide or retinol in cultured whole rat embryos. *Drug Metab Dispos* **20**: 218–225, 1992.
11. Kwart RF, Kimmel CA, Kimmel GL and Slikker W Jr, Identification of the cellular retinoic acid binding protein (CRABP) within the embryonic mouse (CD-1) limb bud. *Teratology* **32**: 103–111, 1985.
12. Howard WB, Sharma RP, Willhite CC and Dawson MI, Binding affinities of retinoids to fetal cellular retinoic acid-binding protein (CRABP) in relation to their teratogenic potency in hamsters. *Biochem Pharmacol* **40**: 643–648, 1990.
13. Apfel C, Crettaz M and LeMotte P, Differential binding and activation of synthetic retinoids to retinoic receptors. In: *Retinoids in Normal Development and Teratogenesis* (Ed. Morriss-Kay G). Oxford University Press, Oxford, in press.
14. Willhite CC, Jurek A, Sharma RP and Dawson MI, Structure-affinity relationships of retinoids with embryonic cellular retinoic acid-binding protein. *Toxicol Appl Pharmacol* **112**: 144–153, 1992.

### Effects of 1,25-dihydroxyvitamin D<sub>3</sub> analogue 1,24(OH)<sub>2</sub>-22-ene-24-cyclopropyl D<sub>3</sub> on proliferation and differentiation of a human megakaryoblastic leukemia cell line

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**Abstract**—The novel analogue of 1,25-dihydroxyvitamin D<sub>3</sub> (1,25(OH)<sub>2</sub> D<sub>3</sub>), 1,24(OH)<sub>2</sub>-22-ene-24-cyclopropyl D<sub>3</sub> (calcipotriol, MC903), exhibits similar effects on cell proliferation and cell differentiation in a newly established human megakaryoblastic leukemia cell line (HIMeg). MC903 was found to inhibit cell proliferation and induce cell differentiation in a liquid culture system at concentrations comparable to those of 1,25(OH)<sub>2</sub> D<sub>3</sub>. Colony formation assay showed that MC903 or 1,25(OH)<sub>2</sub> D<sub>3</sub> markedly diminished the colony-forming ability of HIMeg cells at concentrations of 10<sup>-6</sup> M to 10<sup>-10</sup> M. Cell cycle analysis demonstrated that, as seen with 1,25(OH)<sub>2</sub> D<sub>3</sub>, MC903 also altered the cell cycle distribution; the fraction of cells in G<sub>0</sub> + G<sub>1</sub> increased while those in S and G<sub>2</sub> + M decreased. It can be concluded from these findings that 1,25(OH)<sub>2</sub> D<sub>3</sub> and its analogue MC903 have approximately equipotent effects on cells of megakaryoblastic lineage and are potentially useful in studying the cellular processes that are responsible for megakaryocytopoiesis.

Vitamin D<sub>3</sub> is a secosteroid; sequential hydroxylation in the liver and kidney converts it to 1,25-dihydroxyvitamin D<sub>3</sub> [1,25(OH)<sub>2</sub> D<sub>3</sub>\*] which is the active form. 1,25(OH)<sub>2</sub> D<sub>3</sub> has profound effects on the regulation of bone and mineral metabolism [1]. Most, if not all, of its actions are mediated like steroid hormones via binding and activation

of high affinity nuclear receptors. The activated receptors then modulate gene expression by binding to hormone response elements of the target genes [2]. In classical target tissues, 1,25(OH)<sub>2</sub> D<sub>3</sub> enhances the expression of the calcium-binding protein, alkaline phosphatase, and the 24-hydroxylase [1–3]. Extremely sensitive radioligand receptor binding assays have shown that 1,25(OH)<sub>2</sub> D<sub>3</sub> receptors are present not only in the classical target tissues such as bone, intestinal mucosa and kidney, but also in the skin, endocrine glands, thymus, brain, bone marrow, and activated or transformed lymphopoietic cells, among other tissues [4, 5]. The presence of high affinity 1,25(OH)<sub>2</sub> D<sub>3</sub>

\* Abbreviations: MC903, calcipotriol, 1,24(OH)<sub>2</sub>-22-ene-24-cyclopropyl D<sub>3</sub>; 1,25(OH)<sub>2</sub> D<sub>3</sub>, 1,25-dihydroxyvitamin D<sub>3</sub>; HIMeg, human megakaryoblastic leukemia cell line; IMDM, Iscove's modified Dulbecco's medium; FCS, fetal calf serum.

receptors in so many cell types suggests that the biological functions of this hormone may go well beyond calcium homeostasis; indeed, it has been proposed that  $1,25(\text{OH})_2\text{D}_3$  may be involved in the regulation of gene expression, cell proliferation and cell differentiation in many diverse systems [5–7].

Previous studies showed that  $1,25(\text{OH})_2\text{D}_3$  causes cells from both the murine myeloid leukemia line (M1) and the human promyelocytic leukemia line (HL60) to differentiate to monocyte/macrophage-like cells [4, 8]. Further studies showed that the clonal proliferation of a variety of human acute myelogenous leukemia cell lines is inhibited by  $1,25(\text{OH})_2\text{D}_3$  and this inhibition is independent of the ability of the hormone to induce differentiation of these cells [9]. Clonogenic blast cells from patients with acute myelogenous leukemia are also inhibited in their proliferation by  $1,25(\text{OH})_2\text{D}_3$  [10]. Studies *in vivo* suggest that  $1,25(\text{OH})_2\text{D}_3$  can prolong the survival of mice injected with leukemic cells [8]. The effects of  $1,25(\text{OH})_2\text{D}_3$  on the megakaryocytic lineage are unclear. Vitamin  $\text{D}_3$  metabolites may thus be of potential interest in the treatment of a number of proliferative diseases, but their useful therapeutic range is limited by their potent effects on calcium metabolism, leading to hypercalcemia, hypercalciuria and calcifications in kidney, heart and blood vessels. These considerations have stimulated interest in the development of new vitamin  $\text{D}_3$  derivatives and analogues which retain their regulatory effects on cell proliferation and differentiation but exhibit decreased activity toward calcium metabolism.

MC903 [ $1,24(\text{OH})_2$ -22-ene-24-cyclopropyl  $\text{D}_3$ , calcipotriol] is a cyclopropyl side chain analogue of  $1,25(\text{OH})_2\text{D}_3$  which exhibits similar effects on cell proliferation and differentiation but possesses greatly reduced intestinal calcium absorption and bone calcium mobilization activity [11]. In the present study the comparative effects of MC903 and  $1,25(\text{OH})_2\text{D}_3$  on the proliferation and differentiation of a newly established human megakaryoblastic leukemia cell line (HIMeg) were studied. Our results demonstrate that  $1,25(\text{OH})_2\text{D}_3$  and its analogue have *in vitro* effects on megakaryocytopoiesis.

#### Materials and Methods

**Materials.** MC903 and  $1,25(\text{OH})_2\text{D}_3$  were kind gifts from Dr L. Binderup, Leo Pharmaceutical Products (Ballerup, Denmark). Both compounds were dissolved in

ethanol and stored at  $-20^\circ$ . Iscove's modified Dulbecco's medium (IMDM) powder was from Gibco (Grand Island, NY, U.S.A.). Linbro tissue culture plates (24 well) were obtained from Flow Laboratories (McLean, VA, U.S.A.). All other chemicals and reagents were of analytical grade and obtained commercially.

**Cells and cell culture.** HIMeg cell line was established from a long term culture of peripheral blood cells of a patient with chronic granulocytic leukemia in chronic phase, and it has megakaryoblastic characteristics with ph' chromosome [12]. HIMeg cells were cultured in IMDM medium supplemented with 10% fetal calf serum (FCS), 2 mM L-glutamine, penicillin G (100 U/mL) and streptomycin (100  $\mu\text{g}/\text{mL}$ ) in a humid atmosphere of 5%  $\text{CO}_2$  in air at  $37^\circ$ . One half of the medium was replaced every 3 or 4 days.

**Treatment of cells with hormones.** Cells were plated onto 24-well clusters at cell densities of  $1 \times 10^5$  cells/well in 1.0 mL of IMDM. The test substance,  $1,25(\text{OH})_2\text{D}_3$  or MC903, was added at various concentrations at the beginning of the culture. Equal amounts of ethanol were added to control cultures. The final ethanol concentration was 0.1%. Cells grown under various conditions were harvested and counted and the viability of cells was determined by Trypan blue dye exclusion. Following treatment with MC903 or  $1,25(\text{OH})_2\text{D}_3$ , the morphological characteristics of the cells were determined by examination of Wright-Giemsa-stained cells and multinucleated cells (cells with two or more nuclei) were counted.

**Colony assay.** The colony assay was performed according to the methylcellulose culture method [13]. The HIMeg cells were suspended in medium containing 0.8% methylcellulose (Dow Chemicals, Midland, MI, U.S.A.), 20% FCS and various concentrations of MC903 or  $1,25(\text{OH})_2\text{D}_3$ . Aliquots of culture medium were plated onto 96-well plates at 500 cells/well in 0.1 mL and cultured at  $37^\circ$  in a 5%  $\text{CO}_2$  humidified atmosphere. Incubated plates were cultured for 7 days and were counted with an inverted microscope. A colony formed by HIMeg cells was defined as a cell cluster containing more than 50 cells.

**Flow cytometry.** Cells in log phase of growth were exposed to vitamin  $\text{D}_3$  compounds at  $10^{-6}$  M. Cells were removed from culture at 4 days following exposure, washed with ice-cold phosphate-buffered saline and fixed with ice-cold 70% ethanol. Before analysis, cells were treated with

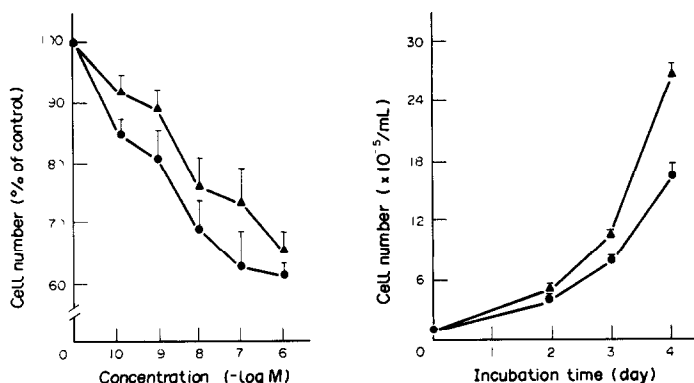


Fig. 1. Effects of MC903 and  $1,25(\text{OH})_2\text{D}_3$  on growth of HIMeg cells. HIMeg cells were plated at  $1 \times 10^5$  cells/mL onto 24-well clusters and incubated in the presence or absence of MC903 (circle) or  $1,25(\text{OH})_2\text{D}_3$  (triangle) at the indicated concentrations for 4 days (left). For time-course studies (right), cells were incubated in the absence (triangle) or presence of  $10^{-7}$  M MC903 (circle) for 0–4 days. Viable cells were counted by Trypan blue dye exclusion. Values represent the means  $\pm$  SEM of four determinations.

RNase A (200  $\mu\text{g/mL}$ ) to reduce fluorescence from RNA and then stained with propidium iodide (10  $\mu\text{g/mL}$ ). DNA-bound propidium iodide was excited at a wavelength of 488 nm and cellular DNA content was determined by fluorescence at a wavelength of 630 nm. Cell cycle distribution was calculated.

### Results

It has been reported that  $1,25(\text{OH})_2 \text{D}_3$  can inhibit the proliferation of leukemia cell lines such as HL-60 and  $\text{U}_{937}$  [4–11]. Therefore, the proliferation and differentiation response of the newly established HIMEg to  $1,25(\text{OH})_2 \text{D}_3$  and MC903 treatment was determined. HIMEg cells were treated with increasing concentrations of MC903 or  $1,25(\text{OH})_2 \text{D}_3$  for 4 days and the total viable cell number was determined. Figure 1 shows that treatment of HIMEg cells with vitamin  $\text{D}_3$  compounds inhibited HIMEg cell proliferation in a dose- and time-dependent manner.

During treatment of HIMEg cells with MC903 or  $1,25(\text{OH})_2 \text{D}_3$ , the morphology of the cells changed significantly from the second day: the cytoplasmic membranes became ruffled, sending out pseudopod-like processes. Multinucleated cells increased in number 4 days after exposure to MC903 or  $1,25(\text{OH})_2 \text{D}_3$  in a dose-dependent manner (Fig. 2). The correlation between these morphological changes and the induction of differentiated characteristics in these cells has been determined [14].

The effects of MC903 or  $1,25(\text{OH})_2 \text{D}_3$  on colony formation by HIMEg cells were investigated with the methylcellulose culture method. As shown in Fig. 3, almost no colony formation was observed in the presence of MC903 or  $1,25(\text{OH})_2 \text{D}_3$  at high concentrations ( $10^{-6}$ – $10^{-9} \text{M}$ ); low concentrations of the vitamin  $\text{D}_3$  compounds could also markedly diminish the colony-forming ability of HIMEg cells.

Cell-cycle profiles were determined on HIMEg cells following exposure to MC903 or  $1,25(\text{OH})_2 \text{D}_3$ . As demonstrated in Fig. 4, significant alterations in cell-cycle distribution occurred 4 days after treatment, i.e. there was a significant increase in the percentage of cells in  $\text{G}_0 + \text{G}_1$  phase while the percentage of cells in both S and  $\text{G}_2 + \text{M}$  phases decreased.

### Discussion

A human megakaryoblastic cell line, HIMEg, was established from a patient with chronic granulocytic

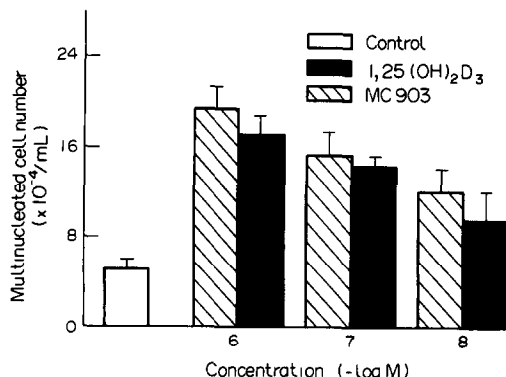


Fig. 2. Effects of MC903 and  $1,25(\text{OH})_2 \text{D}_3$  on the formation of multinucleated cells. HIMEg cells were incubated in the presence or absence of vitamin  $\text{D}_3$  compounds at the indicated concentrations for 4 days and multinucleated cells were counted. Results shown were means  $\pm$  SEM of three determinations. Control denotes untreated cultures. All changes were significantly different from controls ( $P < 0.01$ ).

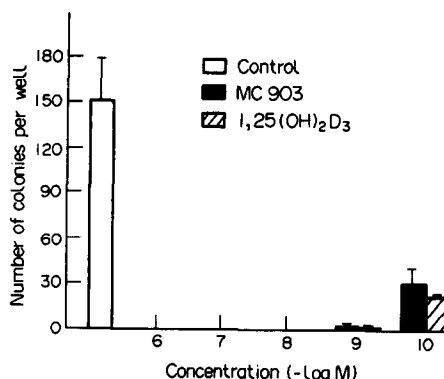


Fig. 3. Effects of MC903 and  $1,25(\text{OH})_2 \text{D}_3$  on colony formation by HIMEg cells. HIMEg cells were plated at 5000 cells/well onto 96-well plates with various concentrations of vitamin  $\text{D}_3$  compounds. After 7 days of culture, colonies were scored under an inverted microscope. Values are means  $\pm$  SEM of six determinations and all changes were significantly different from controls ( $P < 0.001$ ).

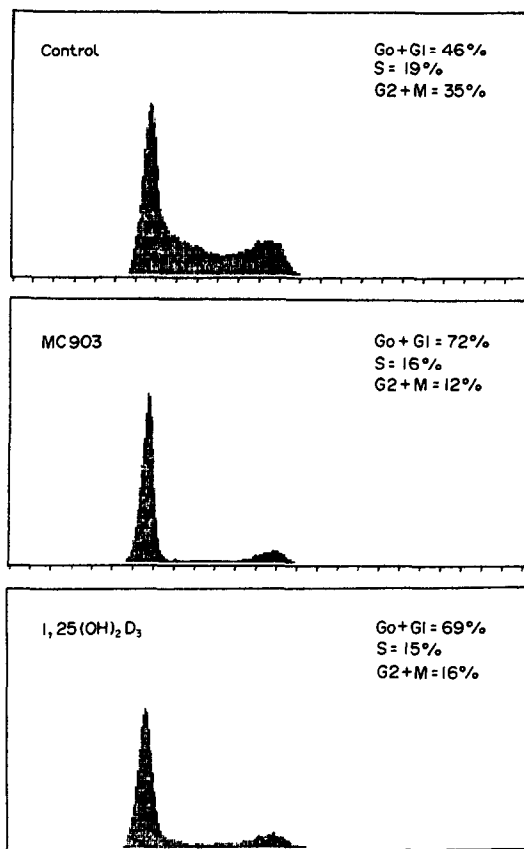


Fig. 4. Cell cycle profiles following exposure to vitamin  $\text{D}_3$  compounds at  $10^{-6} \text{M}$  for 4 days. DNA content of HIMEg cells was determined by fluorescence of propidium bound to DNA. The left-hand peak in the figure represents  $\text{G}_0 + \text{G}_1$  cells (2N DNA), the right-hand peak represents  $\text{G}_2 + \text{M}$  cells (4N DNA) and the area between the two peaks represents cells in S phase (between 2 and 4N DNA). The percentage of cells in these cell-cycle phases is shown in the right part of the figure.

leukemia. HIMEg cells were identified as megakaryoblasts for the following reasons: (a) the platelet peroxidase reaction; (b) the expression of glycoprotein IIb/IIIa (GP IIb/IIIa); and (c) the lack of specific markers for myeloid, monocytic and erythroid lineages [14]. In this report, the comparative effects of  $1,25(\text{OH})_2 \text{D}_3$  and MC903 on the proliferation and differentiation of HIMEg cells were studied. The results show that both  $1,25(\text{OH})_2 \text{D}_3$  and MC903 could significantly inhibit the proliferation and induce the differentiation of HIMEg cells. In a liquid culture system, exposure to MC903 or  $1,25(\text{OH})_2 \text{D}_3$  led to a significant decrease in cell growth rate, and dramatic changes in morphological appearance which were characteristic of differentiation. MC903 and  $1,25(\text{OH})_2 \text{D}_3$  almost completely abolished the colony-forming ability of HIMEg cells at high concentrations ( $10^{-6}$ – $10^{-9}$  M) and markedly diminished the ability at low concentrations ( $10^{-10}$  M). MC903 and  $1,25(\text{OH})_2 \text{D}_3$  were approximately equipotent.

Cell-cycle analysis showed that MC903 and  $1,25(\text{OH})_2 \text{D}_3$  induced significant alterations in cell-cycle distribution. Exposure of HIMEg cells to vitamin  $\text{D}_3$  at  $10^{-6}$  M for 4 days resulted in the accumulation of cells in  $\text{G}_0 + \text{G}_1$  phase while the percentage of cells in S and  $\text{G}_2 + \text{M}$  phases was decreased. This experiment suggests that MC903 and  $1,25(\text{OH})_2 \text{D}_3$  inhibited proliferation of HIMEg cells by slowing transition of the cells from the  $\text{G}_0 + \text{G}_1$  to the S phase, resulting in induction of the  $\text{G}_0 + \text{G}_1$ -arrested cells that differentiate into mature megakaryocytes.

$1,25(\text{OH})_2 \text{D}_3$  is the physiologically important secosteroid that not only mediates calcium reabsorption of bone and intestinal calcium absorption, but also has effects on hematopoiesis [1].  $1,25(\text{OH})_2 \text{D}_3$  induces terminal differentiation of a variety of myeloid leukemic cells from patients and cell lines as well as normal committed myeloid stem cells [4, 10, 15]. The results of the present study further demonstrate that  $1,25(\text{OH})_2 \text{D}_3$  could also play a role in megakaryocytopoiesis; and that MC903, a cyclopropyl side chain analogue of  $1,25(\text{OH})_2 \text{D}_3$ , is as effective as  $1,25(\text{OH})_2 \text{D}_3$  in inhibiting proliferation and stimulating differentiation of HIMEg cells. These effects of  $1,25(\text{OH})_2 \text{D}_3$  are probably mediated directly or indirectly by initial ligand binding to an intracellular  $1,25(\text{OH})_2 \text{D}_3$  receptor, which is a member of the steroid receptor superfamily. Further studies are needed to determine whether there is a functional  $1,25(\text{OH})_2 \text{D}_3$  receptor in HIMEg cells. It will be also necessary to assess the *in vivo* potency of  $1,25(\text{OH})_2 \text{D}_3$  and its derivatives in blocking neoplastic growth.

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## REFERENCES

1. Reichel H, Koeffler HP and Norman AW, The role of the vitamin D endocrine system in health and disease. *N Engl J Med* 320: 980–991, 1989.
2. Haussler MR, Vitamin D receptors: nature and function. *Annu Rev Nutr* 6: 527–563, 1986.
3. Stumpf WE, Sar M and DeLuca HF, In: *Hormonal Control of Calcium Metabolism* (Eds. Cohn DV, Tamage RV and Matthews JL), pp. 222–229. Excerpta Medica, Amsterdam, 1981.
4. Mangelsdorf DI, Koeffler HP, Donaldson CA, Pike JW and Haussler MR,  $1,25$ -Dihydroxyvitamin  $\text{D}_3$  induced differentiation in a human promyelocytic leukemia cell line (HL60): receptor-mediated maturation to macrophage-like cells. *J Cell Biol* 98: 391–398, 1984.
5. Provvedini DM, Tsoukas CD, Deftos LJ and Manolagas SC,  $1,25$ -Dihydroxyvitamin  $\text{D}_3$  receptors in human leukocytes. *Science* 221: 1181–1183, 1983.
6. Dodd R, Cohen M, Newman S and Gray T, Vitamin D metabolites change the phenotype monoblastic U937 cells. *Proc Natl Acad Sci USA* 80: 7538–7541, 1983.
7. Rigby WFC, Denome S and Fanger MW, Regulation of lymphokine production and human T lymphocyte activation by  $1,25$ -dehydroxyvitamin  $\text{D}_3$ . *J Clin Invest* 79: 1659–1665, 1987.
8. Abe E, Miyaura C, Sakagami H, Takada M, Konno K, Yamzaki T, Yoshiki S and Suda T, Differentiation of mouse myeloid leukemia cells induced by  $1,25$ -dihydroxyvitamin  $\text{D}_3$ . *Proc Natl Acad Sci USA* 78: 4990–4994, 1981.
9. Munker R, Norman AW and Koeffler HP, Vitamin D compounds: effect on clonal proliferation and differentiation of human myeloid cells. *J Clin Invest* 78: 424–430, 1986.
10. Koeffler HP, Amatruda T, Ikekawa N, Kobayashi Y and DeLuca HF, Induction of macrophage differentiation of human normal and leukemic myeloid stem cells by  $1,25$ -dihydroxyvitamin  $\text{D}_3$  and its analogues. *Cancer Res* 44: 5624–5628, 1984.
11. Binderup L and Bramm E, Effects of a novel vitamin D analogue MC903 on cell proliferation and differentiation *in vitro* and on calcium metabolism *in vivo*. *Biochem Pharmacol* 37: 889–895, 1988.
12. Wan JH, Mi JX, Qi SL, Wang ZC, Song YH and Yin JL, Establishment of a human megakaryoblastic leukemia cell line and investigation of its biologic characteristics. *Chin J Hematol* 10: 292–294, 1989.
13. Komatsu N, Suda T, Moroi M, Tokuyama N, Sakata Y, Okada M, Nishida T, Hirai Y, Sato T, Fuse A and Miura Y, Growth and differentiation of a human megakaryoblastic cell line, CMK. *Blood* 74: 42–48, 1989.
14. Cheng T, Yan F, Wan JH, Yin JL, Zhang SJ, Qi SL and Song YH, Study on induction of differentiation of a human megakaryoblastic leukemia cell line (HIMEg) by  $1,25$ -dihydroxyvitamin  $\text{D}_3$ . *Sci China [B]* 34: 412–420, 1991.
15. Tanaka H, Abe E, Miyaura C, Kuribayashi T, Konno K, Nishi Y and Suda T,  $1$ -Alpha,  $25$ -dihydroxycholecalciferol and a human myeloid leukemia cell line (HL60). The presence of a cytosol receptor and induction of differentiation. *Biochem J* 204: 713–719, 1982.

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